



The Effect of Bacteria Colony *Pseudomonas fluorescens* (UB_Pf1) and *Bacillus subtilis* (UB_Bs1) on the Mortality of *Pratylenchus coffeae* (Tylenchida: Pratylenchidae)

✉ Presti Mardiyani Purwaningtyas¹, Bambang Tri Rahardjo², Hagus Tarno²

DOI: 10.15294/biosaintifika.v8i3.5067

¹Plant Science Program, Graduate School of Agriculture, Universitas Brawijaya, Malang, Indonesia

²Department of Plant Pests and Diseases, Faculty of Agriculture, Universitas Brawijaya, Malang, Indonesia

History Article

Received 15 February 2016

Approved 30 July 2016

Published 24 December 2016

Keywords:

Bacillus subtilis (UB_Bs1);
Median Lethal Concentration;
Median Lethal Time; *Pseudomonas fluorescens* (UB_Pf1); *Pratylenchus coffeae*

Abstract

Parasitic Root-Lesion nematode of *Pratylenchus coffeae* can reduce the Indonesian coffee plants productivity. Several studies reported that *Pseudomonas fluorescens* and *Bacillus subtilis* endophytic bacteria were antagonistic bacteria to nematode. The objective of this research was to reveal the effectiveness of bacterial colonies density of *P. fluorescens* (UB_Pf1), *B. subtilis* (UB_Bs1), and a combination of both bacteria on nematode mortality using median lethal concentration (LC₅₀) and median lethal time 50 (LT₅₀). The densities of bacteria used in this study were 10⁷, 10⁹, 10¹¹ and 10¹³ cfu/ml. 35 testing nematodes were used and the mortality was counted at 6, 12, 24, 36, and 48 hours after treatments. The results showed that LC₅₀ values of *P. fluorescens* (UB_Pf1) was 4,3x10⁸ cfu/ml, LC₅₀ *B. subtilis* (UB_Bs1) was 1,9x10⁹ cfu/ml, and LC₅₀ combination of both bacteria was, 8x10⁷ cfu/ml. It implies that the application of the combination of both bacteria are more pathogenic than single bacterial treatment. The results also showed that the highest LT₅₀ value was 13.21 hours combination of bacterial colonies with a density of 10¹³ cfu/ml and the lowest LT₅₀ value was 52.00 hours on *P. fluorescens* (UB_Pf1) treatment with colonies density of 10⁷ cfu/ml.

How to Cite

Purwaningtyas, P. M., Rahardjo, B. T., & Tarno, H. (2016). The Effect of Bacteria Colony *Pseudomonas fluorescens* (UB_Pf1) and *Bacillus subtilis* (UB_Bs1) on the Mortality of *Pratylenchus coffeae* (Tylenchida: Pratylenchidae). *Biosaintifika: Journal of Biology & Biology Education*, 8(3), 286-293.

© 2016 Universitas Negeri Semarang

✉ Correspondence Author:
Jl. Veteran, Malang 65145, Indonesia
E-mail: presti_rena@yahoo.co.id

p-ISSN 2085-191X
e-ISSN 2338-7610

INTRODUCTION

Productivity of coffee plants in Indonesia is considered low compared to the main producer countries in the world. One of the causes is the main OPT of parasitic nematode on coffee. The related parasitic nematode that infests almost the whole coffee plantation in Indonesia is the root-lesion nematode of *Pratylenchus coffeae*. The nematode infestation has disturbed the plant growth and production, both quantity and quality.

According to Wiryadi Putra (1998), infestation of *P. coffea* on Robusta coffee could reduce production ranged 28.73%-78.45% or the average of production reduction is about 56.85%. The nematode control has still applied nematicide. Chemical control using nematicide is costly and may create poisonous effect. Besides, greater demand for organic coffee and specialty coffee in the global market, which are safe for human health, are cost higher than the conventional coffee. Therefore, an environmental-friendly method is required as an alternative controlling method. Bacteria are one of organism found abundantly in the soil and has a great potential as bioagent to control nematode, as shown by several genus such as *Pasteuria*, *Pseudomonas*, and *Bacillus* (Emmert & Handelsman, 1999; Siddiqui & Mahmood, 1999).

Bacteria from *Bacillus* spp. And *Pseudomonas* spp. genus are antagonist to nematode and have the greatest population in rhizosphere (Krebs et al., 1998). Thus, this research aimed to reveal LC₅₀ and LT₅₀ value of bacteria collection from Plant Pest and Disease Department, Brawijaya University, *Pseudomonas fluorescens* (UB_Pf1) and *Bacillus subtilis* (UB_Bs1) on *P. coffeae*. Furthermore, the result of this research can be used as references to conduct other researches.

METHODS

This research was conducted at Nematology Laboratory, Plant Pest and Disease Department, Faculty of Agriculture, University of Brawijaya, from January to March 2015. The equipments used were *Baermann's* funnel, *Baermann's* funnel rack, host tweezers, nematode strainer, petri dish, pincers, fishhook needle, micropipette, microscope, hand counter, calibrated beaker, hand sprayer, and other supporting tools. Materials of the research included nematode isolates of *P. coffeae*; bacterial isolate of *P. fluorescens* (UB_Pf1) and *B. subtilis* (UB_Bs1).

Nematode exploration was done through extraction on coffee's roots infected by nematode using *Baermann's* funnel method. The infected

roots, which were infested by *P. coffeae* nematode, were rinsed thoroughly. Then, they were cut into pieces before they were put on the screening paper that had been previously watered until the whole roots submerged for 2 x 24 hours. When suspension containing nematode was obtained, then it was observed using microscope. Lastly, the discovered nematode was hooked in order to be identified by Morphometry technique.

The bacterial isolates used in this research were *P. fluorescens* (UB_Pf1) and *B. subtilis* (UB_Bs1), collections from Plant Pest and Disease Department, Faculty of Agriculture, Brawijaya University. Propagation of both bacteria was done using sterile aqua. The formed bacterial colonies were suspended into sterile water and then they were shook for 24 hours using shaker in order to obtain appropriate concentration of the bacterial colonies density. Several density treatments used were 10⁷, 10⁹, 10¹¹, and 10¹³ cfu/ml. Measurement of the absorbent values used spectrophotometer (OD₆₀₀=1).

The research was conducted by applying appropriate bacterial suspension with a given concentration in a petri dish, which had been inoculated with 35 infective juvenile *P. coffeae*. The observation on mortality of *P. coffeae* was done at 6, 12, 24, 36, and 48 hours after the treatment (hat). The nematode mortality was counted, and then counting the percentage of its mortality. If the control treatment found the mortality less than 20%, the corrected mortality must be counted.

Median Lethal Concentration (LC₅₀) is concentration of bacterial colonies density that causes the nematode mortality reached 50% based on the statistical calculation. While, Median Lethal Time (LT₅₀) is duration of exposure required by bacteria to reach 50% nematode mortality, which is determined by probit analysis method using a software of Hsin Chi Probit Analysis program (Chi, 1997).

RESULTS AND DISCUSSIONS

The result of nematode identification roots of coffee plant

The objective of the identification process in this research was to confirm the testing nematode, *P. coffeae*. The results of identification using the morphometry were showed in the following Figure 1, 2, 3.

Based on the identification, the results showed that the parasitic nematode on the coffee roots included in *P. coffeae*. It could be found from morphometry results and then compared them to the literatures. According to Loof (1949), *P. cof*

feae is marked by two annuls on the labium area, body length 0.37-0.83 mm, the anterior part is dome-shaped. Also, there is a spermatheca that forms a circle and long stylet (14µm-18µm).

According to Siddiqui (1972), female *P. coffeae* has slim shape on younger juvenile and fatter shape on the mature ones. Nematode's body shows noticeable annulations. In general, the lateral part is divided into four sections, but some

of them may be divided into five or six sections. The mouth part has two noticeable annulations. On young female, length of the tail is 2-2.5 times of the body width or 1.5-2 times on the mature female.

If it is compared to some nematode photographs, Mullin (2000) suggested that the nematode also has some similarities in body shape, stylet, basal knob, and the tail. Therefore, it can

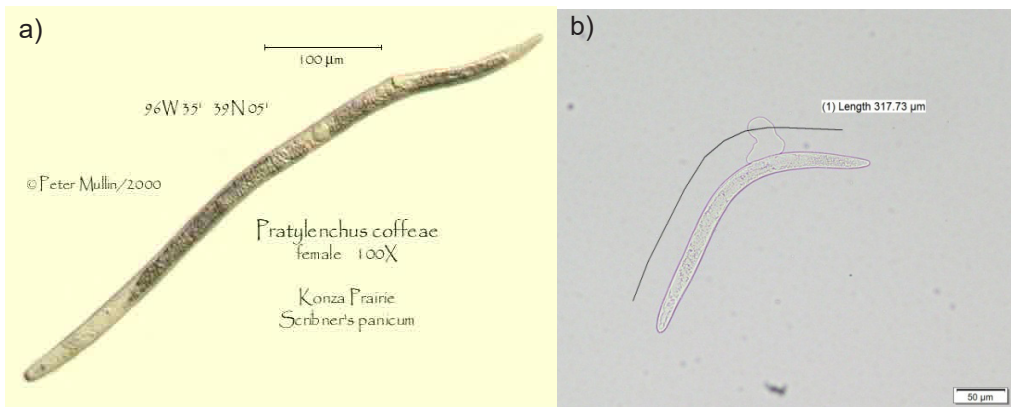


Figure 1. a) *P. coffeae* nematode (Mullin, 2000); b) *P. Coffeae* nematode found at the coffe plant root (100x magnification)

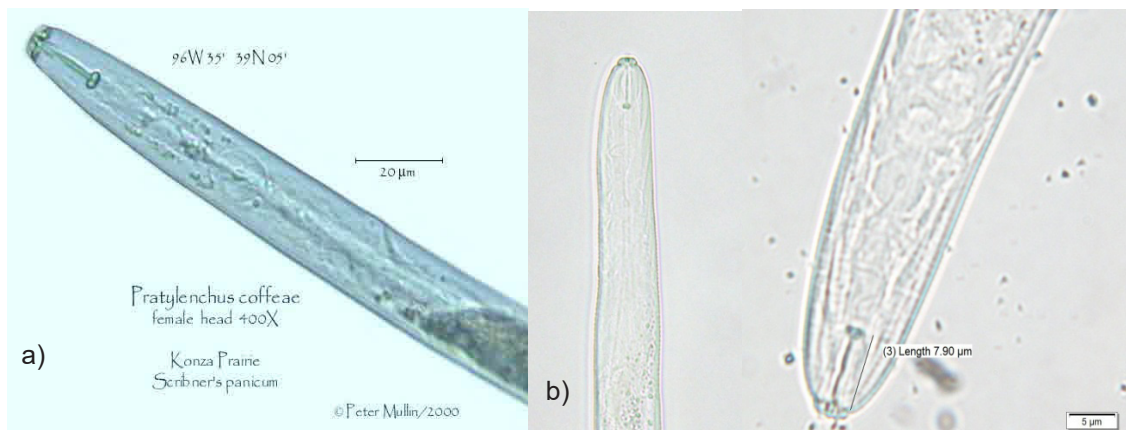


Figure 2. a) *P. coffeae* stylet (Mullin, 2000); b) Nematode stylet (400x magnification)

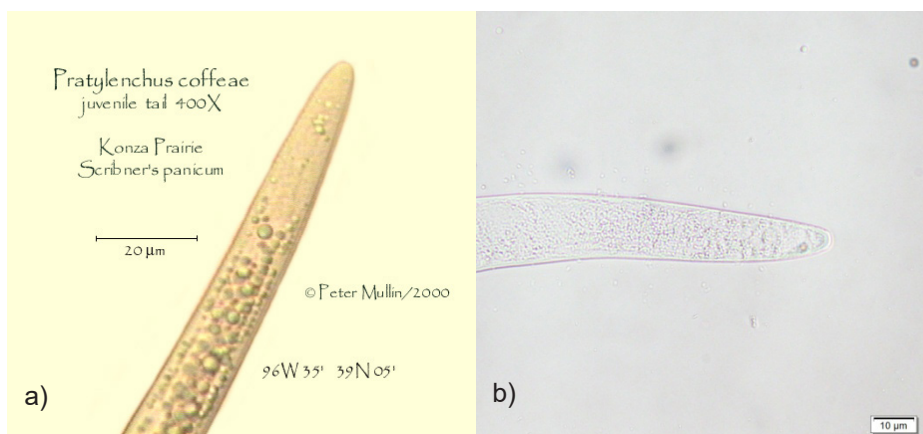


Figure 3. a) *P. coffeae* tail (Mullin, 2000); b) Nematode (400x magnification)

be concluded that the nematode found included in *P. coffeae* species.

The effect of bacterial isolates on percentage of nematode mortality

Based on the results of the analysis of variance test, percentage of nematode mortality showed significant differences on all levels of bacterial colonies density and all intervals of time during observations. Mean percentage of the corrected mortality of nematode on each treatment and time interval of observation were presented in Table 1.

The percentage of the corrected mortality of testing nematode, as presented in Table 1, can be clearly seen in Figure 4. The graphic showed that the testing nematode mortality trend increases along with the length of observation duration. Table 1 and Figure 4 showed that a significant effect of bacterial application on nematode mortality occurred at the entire time of observation started from the first observation (6 hours after application). During the observation, several levels of the applied bacterial colonies density showed a significant effect among treatments. Percentage of the highest mortality was found on the application of both bacterial combinations with the colonies density 10¹¹ cfu/ ml (23.81%), while the percentage of the lowest mortality was found on the application of bacteria *B. subtilis* with colonies density 10⁷ cfu/ml (4.72%).

The same thing was occurred on all observation intervals; the highest mortality occurred on application of both bacterial combination with colonies density 10¹³ cfu/ ml and the percen-

tage of the lowest mortality was occurred on the application of bacteria *B. subtilis* with colonies density 10⁷ cfu/ ml. Yet, for 24 hours observation after the treatment, UB_Pf1 treatment with colonies density 10⁷ showed the lowest mortality. In all of observation time, they showed the increasing percentage of mortality, which meant that the bacterial application affect nematode mortality instantly.

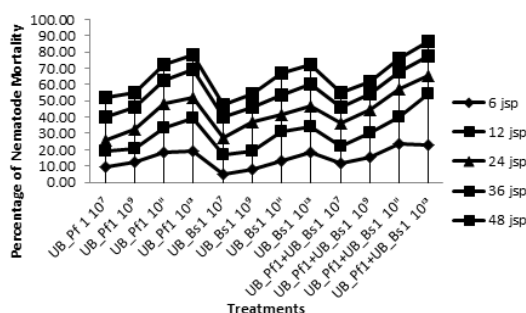


Figure 4. Correlation between concentration of the bacterial colonies density and the nematode mortality

During 36 hours treatment, it was found that UB_Pf1 treatment with the colonies densities 10⁷ and 10⁹, UB_Bs1 with colonies densities 10⁷ and 10⁹, as well as the combination with colonies densities 10⁷ did not show significant differences. The increasing percentages of mortality still kept going on till the last observation (48 hours after the application) and showed high percentage reaching almost 90%. It implies that the bacteria can suppress population of the nematode.

In general, it was found out that bacteria

Table 1. Mean percentage of corrected mortality (%) *P. coffeae* at laboratory

Treatment	Corrected Mortality (%)				
	6 haa ($\bar{x} \pm SD$)	12 haa ($\bar{x} \pm SD$)	24 haa ($\bar{x} \pm SD$)	36 haa ($\bar{x} \pm SD$)	48 haa ($\bar{x} \pm SD$)
Control	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	3.33 ± 1.97	5.33a ± 0.59
UB_Pf1 10 ⁷	9.52 ± 0.74 c	19.05 ± 0.55 ab	25.71 ± 0.82 a	33.75 ± 1.35 a	43.91 ± 4.53 ab
UB_Pf1 10 ⁹	12.38 ± 0.65 d	20.95 ± 1.06 ab	32.38 ± 0.86 b	39.94 ± 2.65 a	47.20 ± 0.44 bc
UB_Pf1 10 ¹¹	18.10 ± 0.55 f	33.33 ± 0.42 c	48.57 ± 0.62 ef	58.71 ± 3.58 cd	67.32 ± 3.82 ef
UB_Pf1 10 ¹³	19.05 ± 0.55 f	39.05 ± 1.41 de	52.38 ± 1.52 fg	66.38f ± 2.80 e	74.18 ± 3.62 f
UB_Bs1 10 ⁷	4.72 ± 1.13 a	17.14 ± 0.99 a	27.62 ± 0.46 a	33.48 ± 3.21 a	38.12 ± 3.09 a
UB_Bs1 10 ⁹	7.62 ± 0.87 b	19.05 ± 0.55 ab	37.14 ± 1.19 bc	39.80 ± 2.93 a	45.98 ± 3.29 ab
UB_Bs1 10 ¹¹	13.33 ± 0.65 de	31.43 ± 0.75 c	41.91 ± 0.02 cd	48.25 ± 2.69 b	60.58 ± 3.74 de
UB_Bs1 10 ¹³	18.10 ± 0.55 f	34.29 ± 1.44 cd	46.67 ± 0.96 def	55.80 ± 1.11 bc	67.47 ± 2.31 ef
UB_Pf1+UB_Bs1(10 ⁷)	11.43 ± 0.00 cd	21.91 ± 0.51 b	36.19 ± 0.41 b	39.87 ± 2.31 a	47.24 ± 1.98 bc
UB_Pf1+UB_Bs1(10 ⁹)	15.24 ± 0.60 e	30.48 ± 1.81 c	44.76 ± 0.99 de	49.29 ± 2.64 b	55.02 ± 2.07 cd
UB_Pf1+UB_Bs1(10 ¹¹)	23.81 ± 0.48 g	40.00 ± 0.67 e	57.14 ± 0.59 g	64.17 ± 1.08 de	71.88 ± 1.30 f
UB_Pf1+UB_Bs1(10 ¹³)	22.86 ± 0.86 g	54.29 ± 0.60 f	65.71 ± 1.11 h	74.84 ± 3.05 f	84.29 ± 1.08 g

Notes :

HAA: Hour After Application

Numbers followed by the same letter in the same column show insignificant difference based on Duncan test at level 5%; data is transformed by equation arcsin \sqrt{x} for statistical analysis purpose.

that gave the greatest effect on nematode mortality were the combination of *P. fluorescens* and *B. Subtilis* with the highest density 10^{13} cfu/ ml. Application that had the lowest percentage was the application of *B. subtilis* with density 10^7 cfu/ml. It reveals that both bacteria are compatible when they are combined, so that their ability in suppressing population of nematode also increases. In line with Jetiyanon & Kloepper's (2002) research suggested that the application of bacteria by combining several compatible bacterial strains and their antagonistic mechanism will provide greater suppressing effect in comparison with single application. Furthermore, the results of the test show that higher colonies density applied will be greater and faster in affecting nematode mortality.

The effect of the bacterial isolates on median Lethal Concentration (LC₅₀) *P. coffeae*

Based on the result of laboratory test, it showed that each bacterium in those treatments had diverse value of LC₅₀. LC₅₀ value, regression equation, and R² of each treatment were presented in Table 2.

Data in Table 2 presented LC₅₀ value on bacterial application of *B. subtilis* (UB_Bs1) $1,9 \times 10^9$ cfu/ ml. It meant that the bacterium was able to cause nematode mortality for almost 50% in 48 hours by applying bacteria that had colonies density 1.9×10^9 cfu/ ml. The results of data analysis showed that the regression equation on the treatment was $y = 3.889 + 0.120x$. It indicated that each increasing coefficient value x indicating the concentration, the coefficient value indicating the probit would increase 3.889. Testing on bacterium *P. fluorescens* (UB_Pf1) had 4.3×10^8 cfu/ ml LC₅₀ value. The value implied that the bacterial colonies density as required by *P. fluorescens* (UB_Pf1) to exterminate nematode 50% in the same time was lower than the treatment *B. subtilis* (UB_Bs1). Value of the regression equation of UB_Bs1 was $y = 3.747 + 0.145x$. It indicated that each increasing coefficient value x (concentration), the coefficient value y (probit) would increase 3.747.

LC₅₀ value on the combination of *P. fluorescens* (UB_Pf1) and *B. subtilis* (UB_Bs1) was 8.8×10^7 . The result indicated that the combined bacterial application caused nematode mortality

up to 50% in 48 hours by applying bacteria having colonies density 8.8×10^7 cfu/ ml. It implied that if the bacteria were, in the same time, applied in combination, they required lower colonies density compared to single application. Based on result of data analysis, the regression equation was $y = 3.463 + 0.193x$. It indicated that each increasing coefficient value x indicating the concentration, the coefficient value indicating the probit would also increase 3.463. Lower colonies density required might occur due to compatibility exists between both bacteria, so that it can increase their antagonistic to nematode, which is influential to the decreasing numbers of colonies density must be applied.

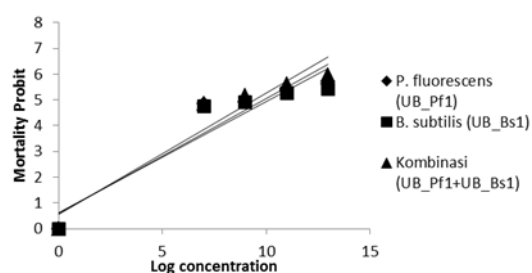


Figure 5. Correlation between concentration of the bacterial colonies density and the nematode mortality

Figure 5 showed the correlation between concentration of bacterial colonies density and testing nematode mortality. Based on the graphic, it presented that the bacterial colonies density had positive correlation on mortality which implied that the increasing bacterial colonies density would be followed by the increasing nematode mortality. It occurs due to the increasing colonies densities that leads to the antagonistic accumulation of the bacteria.

Bacillus is one of the bacterial genres that could inhibit the spread of nematode (Kloepper & Ryu, 2006). The same thing happened in bacteria of Pseudomonas groups, Bacillus has also produce secondary metabolites, and one of them is cytinase enzyme that could degrade the nematode cells. Research conducted by Harni, et al. (2010) reported that the bacterial endophyte isolates from *Bacillus* sp. genus is highly capable in suppressing population of nematode that cause root-lesion, *P. brachyurus*, on patchouli.

Table 2. LC₅₀ value on *P. coffeae* nematode mortality

Bacterial type	LC ₅₀ value	Regression equation	R ²
<i>P. Fluorescens</i> (UB_Pf1)	4.3×10^8 cfu/ml	$y = 3.747 + 0.145x$	0.896
<i>B. subtilis</i> (UB_Bs1)	1.9×10^9 cfu/ml	$y = 3.889 + 0.120x$	0.884
Combination (UB_Pf1 + UB_Bs1)	8.8×10^7 cfu/ml	$y = 3.463 + 0.193x$	0.919

Patil, et al. (2000) stated that chitinolytic enzyme on bacteria is an extracellular enzyme, which takes nutrients and parasitism in controlling diseases biologically. Bacteria from *Aeromonas*, *Alteromonas*, *Chromobacterium*, *Enterobacter*, *Ewingella*, *Pseudoalteromonas*, *Pseudomonas*, *Serratia*, and *Vibriogenuses* are well-known bacteria that have chitinase (Gooday, 1994), *Bacillus* and *Pyrococcus* (Harman et al., 1993).

The effect of bacterial isolates on median Lethal Time (LT₅₀) *P. coffeae*

The result of the probit analysis which was done to find out LT₅₀ showed different values for each treatment. LT₅₀ value and regression equation of each treatment was presented in Table 3.

Data in Table 3 showed that LT₅₀ value by the treatment UB_Pf1+UB_Bs1 10¹³ was 13.211 hours. It indicated that combination of *P. fluorescens* and *B. subtilis* with colonies density 10¹³ cfu/ml caused mortality to the testing nematode up to 50% within 13.211 hours. It implied that it required shorter time than the other treatments. The combination treatment of both bacteria, LT₅₀ value by the treatment UB_Pf1+UB_Bs1 10⁷ was 41.173 hours. It implied that combination of *P. fluorescens* and *B. subtilis* with colonies density 10⁷ cfu/ml required 41.173 hours or 27.962 hours longer than the treatment with colonies density 10¹³ that caused 50% mortality of the testing nematode. Figure 6 presented the regression graphic on bacterial combination treatment of *P. fluorescens* and *B. subtilis* presenting that higher concentration leads to higher value of LT₅₀.

The result of probit analysis on the application of *P. fluorescens* (UB_Pf1) and *B. subtilis* (UB_Bs1) also showed the same results as the combined treatments; the higher concentration of the colonies density applied, the shorter required

time of 50% nematode mortality. It was proven on the application of *P. fluorescens*, the treatment of UB_Pf1 10¹³ had LT₅₀ value 18.900 hours and UB_Pf1 10⁷ with the lowest colonies density had LT₅₀ 52.001 hours. As well as *B. subtilis*, UB_Bs1 10¹³ had LT₅₀ 23.861 hours and LT₅₀ value with UB_Bs1 10⁷ was 50.994 hours.

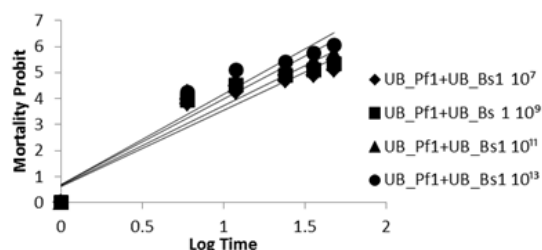


Figure 6. Correlation between time and nematode mortality (Combination)

Based on the result, it can be concluded that the bacterial colonies density on all treatments and LT₅₀ value have positive correlation, in which higher concentration of the bacterial colonies density would shorten the required time to reach mortality up to 50%. In addition, it implies that higher concentration of colonies densities shows that the bacteria would be more toxic. Besides, shorter time required by the bacteria to cause mortality on the testing nematode shows that the bacteria is more toxic than the application of other bacteria, which requires longer time to cause 50% mortality of testing nematode. The correlation can be clearly seen on regression graphic in Figure 7 and Figure 8.

Pseudomonas of *fluorescens* group is one of bacterial group, which is mostly learnt as bio-agent controller. The bacteria have a combination of effective bio controlling mechanisms. *Pseudomonas* produces some secondary metabolites with antimicrobial activities to other bacteria and

Table 3. LT₅₀ value on *P. coffeae* nematode mortality

Treatments	LT ₅₀ value	Regression equation	R ²
UB_Pf1 10 ⁷	52.001 hours	y = 2.485+1.465x	0.912
UB_Pf1 10 ⁹	43.856 hours	y = 2.590+1.468x	0.910
UB_Pf1 10 ¹¹	22.721 hours	y = 2.791+1.629x	0.915
UB_Pf1 10 ¹³	18.900 hours	y = 2.864+1.674x	0.915
UB_Bs1 10 ⁷	50.994 hours	y = 2.215+1.631x	0.935
UB_Bs1 10 ⁹	40.638 hours	y = 2.411+1.609x	0.943
UB_Bs1 10 ¹¹	28.505 hours	y = 2.786+1.522x	0.910
UB_Bs1 10 ¹³	23.861 hours	y = 2.823+1.580x	0.911
UB_Pf1+ UB_Bs1 10 ⁷	41.173 hours	y = 2.703+1.423x	0.907
UB_Pf1+ UB_Bs1 10 ⁹	28.791 hours	y = 2.854+1.471x	0.907
UB_Pf1+ UB_Bs1 10 ¹¹	17.575 hours	y = 2.997+1.609x	0.909
UB_Pf1+ UB_Bs1 10 ¹³	13.211 hours	y = 2.937+1.840x	0.920

pathogens. Besides, the bacteria could inhibit the pathogen's growth by limiting the use of iron in the soil, which utilizing the resulted siderophore (Duijff et al., 1993; Duffy & Defago, 1999).

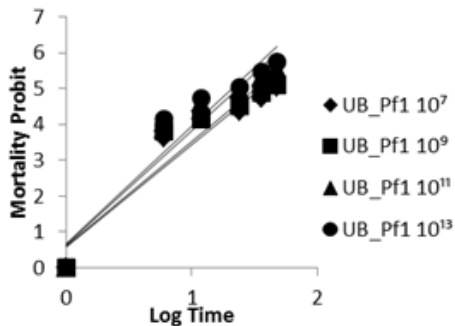


Figure 7. Correlation between time

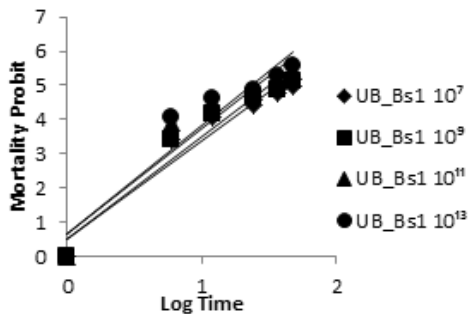


Figure 8. Correlation between time and nematoda mortality (*P. fluorescens*) nematoda mortality (*B. subtilis*)

Gokte & Swarup (1988) reported that *B. subtilis*, *B. cereus*, and *B. pumilus* could inhibit larva-side activities on second juvenile (J2) *Meloidogyne incognita* in vitro. Based on result of the experiment in pot, it showed that reduced development of *M. incognita* in tomato by the application of *B. subtilis*. It is reported that this bacteria are potential to control some parasitic nematodes, including *Meloidogyne* spp.

CONCLUSIONS

The combined treatment of bacterial isolates *P. fluorescens* (Ub_Pf1) and *B. subtilis* (UB_Bs1) provides better results than single application with LC_{50} value (8.8×10^7 cfu/ml). The combined treatment of UB_Pf1+UB_Bs1 with colonies density 10^{13} cfu/ml can kill 50% nematode in the shortest time with LT_{50} value (13.211 hours) and treatment UB_Pf1 with colonies density 10^7 cfu/ml requires the longest killing time of 50% nematode with the LT_{50} value (52.001 hours).

ACKNOWLEDGEMENT

The researchers would like to express their gratitude and great appreciation to the Agricultural Human Resources Development Institute (BPSDMP) and Estate Corps Regional Center of Seed and Protection (BBPPTP) Surabaya for their assistances during this research was conducted.

REFERENCES

- Duffy, B. K. & G. Defago. (1999). Environmental Factors Modulating Antibiotic and Siderophore Biosynthesis by *Pseudomonas fluorescens* Biocontrol Strains. *App. Env. Microb.* 65(6), 2429-2438.
- Duijff, B. J., Meijer, J. W., Bakker, P. A. H. M. & Schippers, B. (1993). Siderophoremediated Competition for Iron and Induced Resistance in The Suppression of Fusarium Wilt of Carnation by Fluorescent *Pseudomonas* spp. *Netherlands Journal of Plant Pathology*, 99(5-6), 277-289.
- Emmert, E. A. B. & Handelsman, J. (1999). Biocontrol of Plant Disease: a (Gram +) Positive Perspective. *FEMS Microbiol Lett*, 171(1), 1-9.
- Gokte, N. & Swarup, G. (1988). On the Potential of Some Bacterial Biocides Against Root-Knot and Cyst Nematodes. *Indian Journal of Nematology*, 18(1), 152-153.
- Gooday, G. W. (1994). Physiology of Microbial Degradation of Chitin and Chitosan. in Ratledge C, editor. *Biochemistry of Microbial Degradation*. Netherlands: Kluwer Academic Publ.
- Harman, G. E., Hayes, C. K., Lorito, M., Broadway, R. M., Di Pietro, A., Peterbauer, C., & Tronsmo, A. (1993). Chitinolytic Enzymes of *Trichoderma harzianum*: Purification of Chitobiosidase and Endochitinase. *Phytopathology*, 83(3), 313-318.
- Harni, R. & Khaerati. (2013). Evaluasi Bakteri endofit untuk Pengendalian Nematoda *Pratylenchus coffeae* pada Tanaman Kopi. *Buletin Ristri*, 4(2), 109-116.
- Jetiyanon, K., & Kloepper, J. W. (2002). Mixtures of Plant Growth-Promoting Rhizobacteria for Induction of Systemic Resistance Againsts Multiple Plant Diseases. *Biol. Control*, 24(3), 285-291.
- Kloepper, J. & Ryu, C. (2006). Bacterial Endophytes as Elicitors of Induced Systemic Resistance. in *Microbial Root Endophytes*, eds. Schulz, B., C. Boyle, T. Siebern. Springer-Verlag, Heidelberg.
- Krebs, B., B., Hoeding, S., Kuebart, M. A., Workie, H., Junge, G., Schmiedeknecht, R., Grosch, H., Bochow, & M. Havesi. (1998). Use of *Bacillus subtilis* as Biocontrol Agent: Activities and Characterization of *Bacillus subtilis* Strain. *Zeitschrift Pflanzenkrankh Pflanzenschutz*, 105, 181-197.
- Loof, P. A. A. (1949). The Family Pratylenchidae .Thorne. in William R Nickle (ed) 1991. *Manual of agriculture Nematology*. Mercel Dekker, Inc New York.
- Mullin, P. (2000). Photo Gallery *Pratylenchus coffeae*

- Konza Prairie. Retrieved from <http://nematode.unl.edu/pracoff.htm>
- Siddiqui, Z. A. & Mahmood, I. (1999). Role of Bacteria in The Management of Plant Parasitic Nematodes (a review). *Bioresource Technol*, 69(2), 167-179.
- Siddiqui, M. R. (1972). *Pratylenchus coffeae* CIH. Description of Plant Parasitic Nematodes Set 1.6. Wallingford UK. CAB International: 3
- Patil, R. S., Ghormade V., & M. V. Deshpande. (2000). Chitinolytic Enzymes: An Exploration. *Enzym and Microbial Technology*, 26(7), 473-483.
- Wiryadi Putra, S. (1998). *Pengelolaan Nematoda Parasit pada Tanaman Kopi di Indonesia*. Kumpulan Materi Pelatihan Pengelolaan Organisme Pengganggu Tanaman Kopi No. Seri 017PL T05.98. Pusat Penelitian Kopi dan Kakao. Indonesian Coffee and Cocoa Research Institute: 1-15.